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Functionalized SERS substrate based on silicon nanowires for rapid detection of prostate specific antigen

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Abstract

Owing to its high sensitivity, surface-enhanced Raman spectroscopy (SERS) is largely used in biosensing as an efficient optical technique. In this work, we report the elaboration of a novel SERS substrate based aptasensor for ultra-sensitive and selective detection of prostate specific antigen (PSA). The SERS substrate is based on silicon nanowires (SiNWs) as transducer surface coated by silver nanoparticles (AgNPs). The silver nanoparticles have two purposes : a) aptamer immobilization via thiol chemistry and b) Raman signal enhancement. The presence of PSA induces the appearance of amide vibration modes only if the aptamer is present on the surface of SiNWs. The platform is highly sensitive, selective and specific to PSA in a wide range of concentrations from 0.1 to 20 $\mu\text{g.L}^{-1}$ with a detection limit of 0.1 $\mu\text{g.L}^{-1}$, which encompasses the blood serum range of healthy subjects and ill patients. This optical biosensor shows a great stability and it is successfully validated in human blood serum samples showing very good results.

Keywords

SERS; SiNWs; AgNPs; PSA; Aptasensor

1. Introduction

Cancer has been ranked on top as one of the major challenges for human survival due to its great threat to health worldwide. Prostate cancer is the most common cancer related to men over 50 years. It has a high mortality rate when it is diagnosed in advanced stages [1-3]. It is recommended that subjects with a family history with this cancer have to check themselves from the age of 45. Early individual diagnosis is urgently needed to reduce mortality, to increase the life expectancy (at least 10 years) and to improve the treatment outcome [4]. Early diagnosis of prostate cancer is related to prostate specific antigen (PSA) that is regarded as a gold standard biomarker. PSA is a 34-KDa single-chain glycoprotein secreted by the prostate gland [3]. The normal level of this protein in the serum is less than $4 \mu\text{g.L}^{-1}$. A level above this value is often suspected when malformation or tumors appear in the prostate gland [5]. To date, several analytical assays such as enzyme-linked immunosorbent assay [6, 7], chemiluminescence immunoassay [8], immunosensors [9], radioimmunoassay [10] and luminescent immunoassay are commonly used to determine PSA levels in blood serum [11].

On another hand, SERS is also a powerful analytical tool that can be used for detection of proteins, DNA and small molecules [12]. Compared with fluorescence and electrochemistry based methods, SERS can provide steady and sharp fingerprint spectra in complex samples with a low background interference [13] thanks to its inherent characteristics such as short analysis time, high sensitivity, low detection limit, small sample volume requirement and its non-destructive nature [14, 15].

Several developed strategies are based on antibody/antigen biorecognition. However, antibodies are subject to degradation and are bulky biomolecules (160 KDa) [16]. In addition, in vivo preparation of antibodies is expensive, time consuming and difficult to achieve [17]. For these reasons, the choice of antibody-based biosensors is being replaced by aptamer-based sensors. In fact, the rapid emergence of aptamers as alternative capture molecules has opened the way to new diagnostic platforms. Aptamers are single-stranded DNA or RNA oligonucleotides selected by systematic evolution of ligands by exponential enrichment (SELEX) [18]. Their interest lies in their high selectivity, specificity, binding affinity and stability, which allow them differentiating isoforms and splicing variants of a protein and to form distinct secondary structures capable of binding to RNA, DNA or target proteins. It is also easy to chemically modify them by many functional groups, such as amine biotin and thiol groups. In addition, aptamers could be denatured and renatured several times [19]. In the last decades, a lot of

work has been done to build new aptamer-based methods for PSA detection such as electrochemical [20, 21], fluorescence [22] and SERS techniques [23].

Another important parameter in biosensing is the transducer material. Silicon nanowires, which are a one-dimensional nanomaterial with a large specific surface area [24], are one of the most used nanomaterial in SERS-based biosensors. Taking advantage of the characteristic surface plasmon resonance of silver nanoparticles [25], which act as signal enhancing substrates, the combination of these two materials has been widely used as a transducer material to develop SERS sensors [26]. In view of the above, we report here the development of a SERS aptasensor for selective, sensitive and inexpensive prostate specific antigen detection. The SERS substrate was functionalized first by a very dense layer of self-assembled hexanethiol, then partially exchanged by the self-assembled thiol-terminated DNA aptamer for target recognition. The platform serves to sensitively and selectively detect PSA in spiked PBS solutions. Upon optimization of the detection parameters, the device was successfully applied to detect PSA in spiked human serum samples.

2. Materials and methods

2.1. Reagents

Silver nitrate, hydrofluoric acid (40%), hydrogen peroxide (34%), nitric acid (70%), sulfuric acid (97%) and hexanethiol (MCH) were purchased from Sigma-Aldrich (www.sigmaaldrich.com). All these solutions were prepared using ultrapure water produced by a Millipore system. Anti-PSA specific DNA aptamer modified with thiol group at the 5' end (5' C₆H₁₃S-TTTTAAATTAAGCTCGCCATCAAATAGCTTT-3') was acquired from Biomers (Germany, www.biomers.net). PSA molecules were purchased from Merck-Millipore (Massachusetts, USA, www.merckmillipore.com).

2.2. Apparatuses

SERS measurements were performed at room temperature (RT) on a Micro Raman HORIBA system (LabRAM HR800). A helium-Neon laser excitation 632.8 nm was used and has a power of about 5 mW at the sample surface.

The morphological properties and the composition analysis of the as prepared samples were assessed by scanning electron microscopy (SEM) using respectively a JEOL JSM 7100F thermal field emission electron gun microscope and a HITACHI FLEX SEM II microscope equipped with an Energy Dispersive of X ray (EDX) detector. The XRD spectra were recorded using a Bruker D8 diffractometer operating with Cu Ka1/Ka2 radiation. The absorbance spec-

tra were measured using UV-Vis-NIR spectrophotometer (Perkin-Elmer Lambda 950) at RT in the wavelength range from 400 to 500 nm.

2.3. Bioplatfrom assembly and operation

The stepwise development of our label-free SERS aptasensing platform for PSA is illustrated in **Fig. 1**. Briefly, SiNWs were prepared by metal-assisted chemical etching (MACE) from lightly N-doped Si (100) wafers [27]. Then, AgNPs were deposited on SiNWs nanostructure through the electroless deposition technique by the reduction of Ag^+ [28]. Then, SiNWs were immersed in an aqueous solution of HF and AgNO_3 under the optimal conditions found in our previous study devoted for rhodamine 6G detection [26]. The AgNPs/SiNWs were further functionalized with a densely packed self-assembled layer of hexanethiol, to avoid non-specific binding of the analyte on the surface, by incubating the SERS substrate in 10^{-3} M of MCH [29, 30]. Since the arrangement of the thiolated DNA on sensor surfaces has a deep impact on molecular recognition [31], the thiol-terminated DNA aptamer (anti-PSA) was tethered to the surface by exchanging with already adsorbed thiols through immersing MCH/AgNPs/SiNWs substrate ($1 \times 1 \text{ cm}^2$ wafer) in 2 mL of 1 μM of anti-PSA aptamer dissolved in PBS for 4, 8, 12 and 16h at RT. The wafer was removed from the solution, washed twice with PBS solution and twice with deionized water then dried under a gentle N_2 flux. In the last step, the aptasensor (anti-PSA/MCH/AgNPs/SiNWs) was incubated for 30 min with different concentrations of PSA dissolved in phosphate buffered saline solution at pH 7.4 [32]. Finally, the prepared substrates were thoroughly washed and dried before Raman measurement.

Figure 1

3. Results and discussion

3.1. Morphological characterization

The SEM cross section image of SiNWs shows a one-dimensional structure with a thickness of about 15 μm as it is displayed in **Fig. 2A**. The SEM surface image of AgNPs/SiNWs presented in **Fig. 2B** shows that the AgNPs have almost spherical shapes with a relatively uniform size (mean diameter of 100 nm) and they are uniformly distributed over the surface of the SiNWs. The EDX analysis have been performed on the SEM cross-sectional of the SiNWs before and after the deposition of the AgNPs. **Fig. 2C** displays the EDX spectrum of AgNPs / SiNWs from the surface to 1.6 μm , this spectrum shows that the mass percentage of silver in

this area is of the order of 2 %. This percentage decreases with depth until it cancels at a depth of 4.5 μm as it can be seen in **Fig. 2D**.

The UV-Vis absorbance spectra of SiNWs and AgNPs/SiNWs show absorbance values above 96 % over a wavelength range from 400 to 500 nm. The AgNPs/SiNWs spectrum is characterized by the presence of a weak plasmon band of AgNPs at about 440 nm [33-34]. This weak band is most likely due to the low density of silver nanoparticles on the surface of SiNWs (**Fig. 2E**).

The XRD spectrum of SiNWs shows the existence of some silicon diffraction peaks at $2\theta = 32^\circ$ and 61.5° which are related to the crystallographic planes of silicon [200] and [004], respectively. The XRD spectrum of AgNPs/SiNWs shows, in addition to the peaks of SiNWs, the existence of other peaks at $2\theta = 37.9^\circ$, 44.3° and 56.2° which are respectively attributed to [111], [200] and [211] planes of the faceted cubic silver crystals [35, 36] as it can be seen in **Fig. 2F**. This spectrum also confirms the presence of the silver nanoparticles on the SiNWs surface.

Figure 2

3.2. Optimization of the aptamer incubation time

The surface modification of MCH/AgNPs/SiNWs by anti-PSA aptamer was evaluated by Raman. As displayed in **Fig. 3A** and **Fig. 3B**, which show the SERS spectra of MCH/AgNPs/SiNWs and anti-PSA/MCH/AgNPs/SiNWs for different incubation times (4, 8, 12 and 16 hours) of the substrate in aptamer, dissolved at $1\mu\text{M}$ in PBS, for both frequency ranges of $550\text{-}1700\text{ cm}^{-1}$ and $2800\text{-}3100\text{ cm}^{-1}$, respectively. The control Raman spectrum of MCH/AgNPs/SiNWs highlights the vibration modes of the adsorbed hexanethiol. The main vibration modes are located at 632 and 706 cm^{-1} and attributable to $\nu(\text{C-S})_{\text{G}}$ and $\nu(\text{C-S})_{\text{T}}$, respectively. Additionally, the modes at $890, 1080, 2867, 2931$ and 3060 cm^{-1} are attributed to $\rho(\text{CH}_3)$, $\nu(\text{C-C})_{\text{G}}$, $\nu(\text{CH}_3)$, $\nu(\text{CH}_2)_{\text{s}}$ and $\nu(\text{CH}_2)_{\text{as}}$, respectively [37-39].

For longer incubation times, several changes are visible. As it is shown in **Fig. 3A** and **Fig. 3B**, MCH modes undergo a decrease in their scattered intensities, such as the modes at 632 , 706 and 1080 cm^{-1} , whereas some others experience an increase such as at 2867 and 2931 cm^{-1} . The decreased vibrations are the eigen modes of MCH while the other ones (CH_2) exist also in aptamer and the increase of their intensities indicates that the amount of CH_2 groups on the surface is becoming higher. **Fig. 3A** shows also the apparition of time-dependent new vibration modes such as the modes at $960, 1107$ and 1449 cm^{-1} , which are the eigen modes at-

tributed to NH₂ group of adenine, PO₂ stretching and deoxyribose, respectively [40-43]. The intensity decrease of MCH vibration modes and concomitant intensity increase of aptamer vibrations can be explained by the exchanging of a densely packed self-assembled layer of hexanethiol by a thiolated anti-PSA aptamer.

3.3. Optimization of the SERS response to the presence of PSA

The PSA signature is closely dependent on the amount of aptamer present in the surface of the SERS substrate. So, the effect of varying the incubation time (4h, 8h, 12h and 16h) of MCH/AgNPs/SiNWs in aptamer solutions was studied to determine the fittest one. **Fig. 3C** shows the SERS spectra of PSA (1 µg.L⁻¹)/anti-PSA/MCH/AgNPs/SiNWs substrate prepared at different incubation times in aptamer solutions. At relatively short incubation times in aptamer (4h and 8h), the PSA/ anti-PSA/MCH/AgNPs/SiNWs spectra do not show clearly the Raman vibration modes of PSA. For times longer than 12 hours, these spectra show clearly the PSA signature, through the Raman vibration modes of amides. For instance, the newly visible peaks are located at 1650, 1550, 1300 cm⁻¹ are respectively assigned to amides I, II and III [44-46]. The intensities of these modes increase with time from 4h to 16h. The incubation time of MCH/AgNPs/SiNWs in aptamer was chosen 16h to carry out the work. Times superior to 16h were not assayed since the latter gives satisfactory results and is a good compromise between overall experiment duration and sensitive detection of the target.

3.4. Sensing of PSA

The **Fig. 3D** presents the SERS spectra of the aptasensing platform before and after the incubation of anti-PSA(16h)/MCH/AgNPs/SiNWs substrate in PSA solutions ranging from 0.1 to 20 µg.L⁻¹. As it can be seen in **this figure**, the addition of PSA solution induces the appearance of new peaks, such as the vibration modes of amide III at 1300 cm⁻¹ and amide II at 1550 cm⁻¹. The intensity of these modes increases concomitantly with the increase of PSA concentration. This increase indicates that the PSA amounts on the surface is becoming larger. Accordingly, the intensities of the Raman vibration mode at 1550 cm⁻¹ were used to plot the calibration curve representing the Raman intensity vs. the target concentration in solution. The aptasensor is able to detect PSA in a wide linear range of concentration ranging from 0.1 to 20 µg.L⁻¹. The linear regression equation, shown in Figure **Fig. 3E**, is:

$$\text{Raman intensity (a.u)} = 26 [\text{PSA}] (\mu\text{g.L}^{-1}) + 300$$

The correlation coefficient of the linear curve is $R^2 = 0.992$ and the obtained limit of detection (LOD) is 0.1 µg.L⁻¹. This parameter was calculated according to the $3S_b/m$ criterion, where m

is the slope of the calibration curve and S_b was estimated as the standard deviation of three different measurements recorded for the lowest analyte concentration measured.

Figure 3

The reported label-free aptasensor for PSA detection shows an excellent analytical behavior compared to previously reported works as displayed in **Table 1**. For instance, it is outperformed by the electrochemical biosensors reported in [32], [37] and [50] in terms of LOD and dynamic ranges but this biosensor is easier to build and have a higher self-life (up to 2 months). Moreover, it performed better than those using fluorescence and SERS spectroscopies reported in references [51] and [52] using polymer quantum dots or two antibodies, respectively. Overall, this device displays satisfactory performances in terms of linear range and limit of detection, which remains sufficient to conveniently detect the analyte in human blood serum with moderate dilutions of the serum samples. Moreover, the use of an anti-PSA aptamer instead of antibodies insure more stability and a low-cost device. The SERS-based techniques used generally complexed methods based on sandwich-type bioassay however our biosensor is based on direct method of detection. It is of paramount importance to mention that this work is the first to use aptamer for PSA detection by the SERS technique.

Table 1

3.5. Performance of the biosensor

Selectivity and specificity are two very important characteristics that define a given aptasensing performance and validity. These two characteristics were evaluated in the presence of human serum albumin (HSA) and bovine serum albumin (BSA) as competing proteins to PSA. The selectivity and specificity tests were performed in the same conditions of PSA detection by adding $10 \mu\text{g.L}^{-1}$ of each interferent.

As it can be seen in **Fig. 4A** and **Fig. 1S** an excellent discrimination was observed in the SERS spectra intensity at 1550 cm^{-1} in presence of $10 \mu\text{g.L}^{-1}$ of the different proteins PSA, HSA and BSA, respectively. PSA gave a clear increase in SERS intensity at 1550 cm^{-1} , which corresponds to amide II, but the other two proteins HSA and BSA do not cause any significant change in the SERS spectra of the biosensor, indicating a very good selectivity towards PSA.

In addition, to illustrate the specificity, the aptasensor was tested by measuring and comparing the response of the two as-mentioned proteins in presence and in absence of PSA. No significant change in the SERS spectra of the proteins was observed without PSA, compared to

those with PSA as it can be shown in **Fig. 4A** and **Fig. 1S**, indicating good specificity of the developed biosensing system.

The storage stability of the aptasensor was checked for a period of two months. The anti-PSA/MCH/AgNPs/SiNWs was kept in refrigerator at 4° C. The SERS responses provided by the aptasensors with and without PSA were recorded each control day. **Fig. 4B** and **Fig. 2S** show that the designed biosensor exhibits a stable signal either in absence or in the presence of 5 $\mu\text{g.L}^{-1}$ of PSA after 62 days of storage.

Figure 4

3.6. Applicability in complex matrix

Since this aptasensor has shown good selectivity against the proteins HSA and BSA existing in human blood serum, the selectivity of this device has also been tested in more complex samples (female blood serum purchased from Sigma-Aldich). It is worth noting that the levels of PSA in female serum blood is lower than 0.1 $\mu\text{g.L}^{-1}$ [55], so it should not interfere in our assay since it below the limit of detection of the biosensor. In this context, two concentrations of PSA, 3 $\mu\text{g.L}^{-1}$ and 6 $\mu\text{g.L}^{-1}$, were added to the serum, which was diluted three times in PBS to reduce its viscosity. The detection process is the same as for a buffer solution. The **Fig. 3S** displays the response SERS of the aptasensor before and after the addition of serum, 3 $\mu\text{g.L}^{-1}$ and 6 $\mu\text{g.L}^{-1}$ of PSA. This figure shows that the vibration modes of amide II at 1550 cm^{-1} appears after the addition of PSA and their intensities increase with increasing PSA concentrations. The analytical results are gathered in **Table 2**. The detected values of the PSA concentrations after serum spiking (F) are close to those added ones (P). The recovery values, presented in **Table 2**, are about 102 and 103% for two samples (1 and 2). These figures show that the method can be reasonably applied for physiological fluids.

Table 2

4. Conclusion

We reported in the design of new bioplatfrom using aptamer tethered to AgNPs/SiNWs SERS substrate to detect selectively with specificity PSA form using an optic technique. The design stepwise procedure is simple yet giving high performance results. The detection of PSA was monitored by the apparition of amide vibration modes and the increasing of their intensities is timely dependant of the PSA concentrations in solution. The developed aptasensor is able to detect PSA in wide linear range of concentrations from 0.1 to 20 $\mu\text{g.L}^{-1}$ in spiked PBS solutions. Finally, we validated in human serum samples as complex matrix and it shows very

satisfactory results despite the complexity of human fluids. The amenable procedure can be generalized to other proteins/aptamers or small molecules/aptamers using SERS or other optical techniques.

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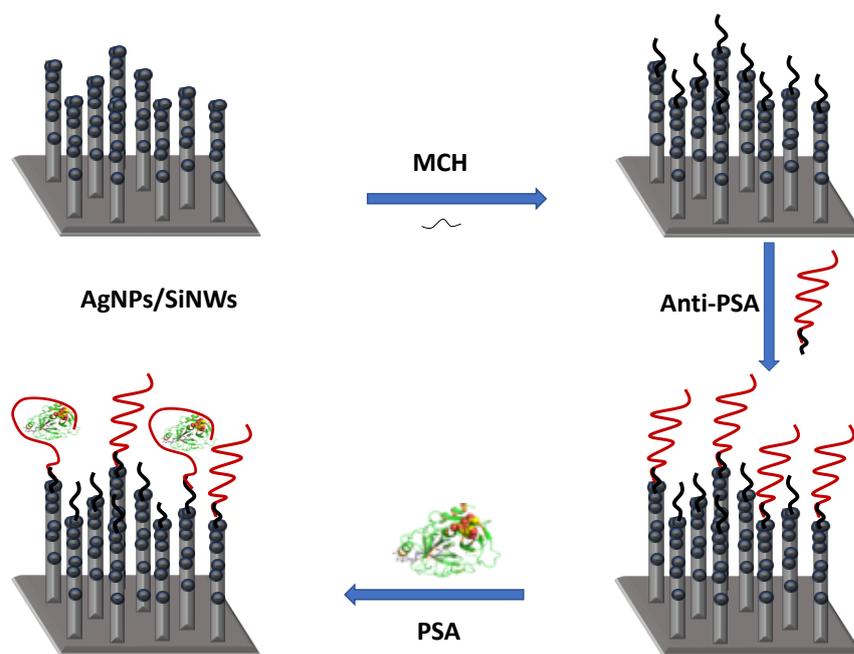


Figure 1. Schematic representation of the strategy of the stepwise assembly of the bioplatfrom and operation.

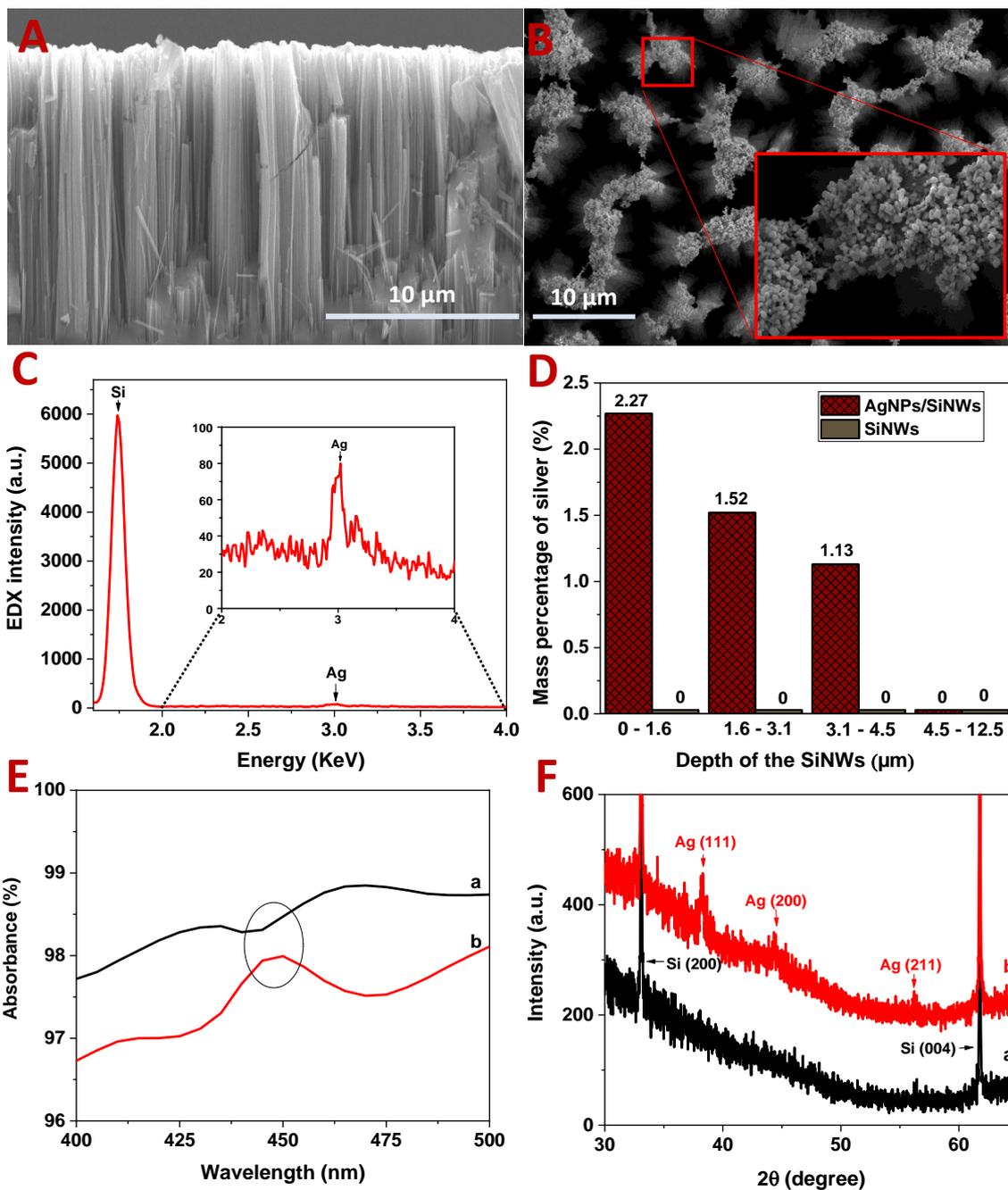


Figure 2. (A) SEM cross-section image of SiNWs, (B) SEM top surface view of AgNPs/SiNWs, (C) EDX spectrum of AgNPs/SiNWs from the surface until 1.6 μm , (D) Mass percentage of silver for SiNWs in depth before and after AgNPs decoration, (E) UV-visible absorbance spectra and (F) XRD patterns and of SiNWs (a) and AgNPs/SiNWs (b).

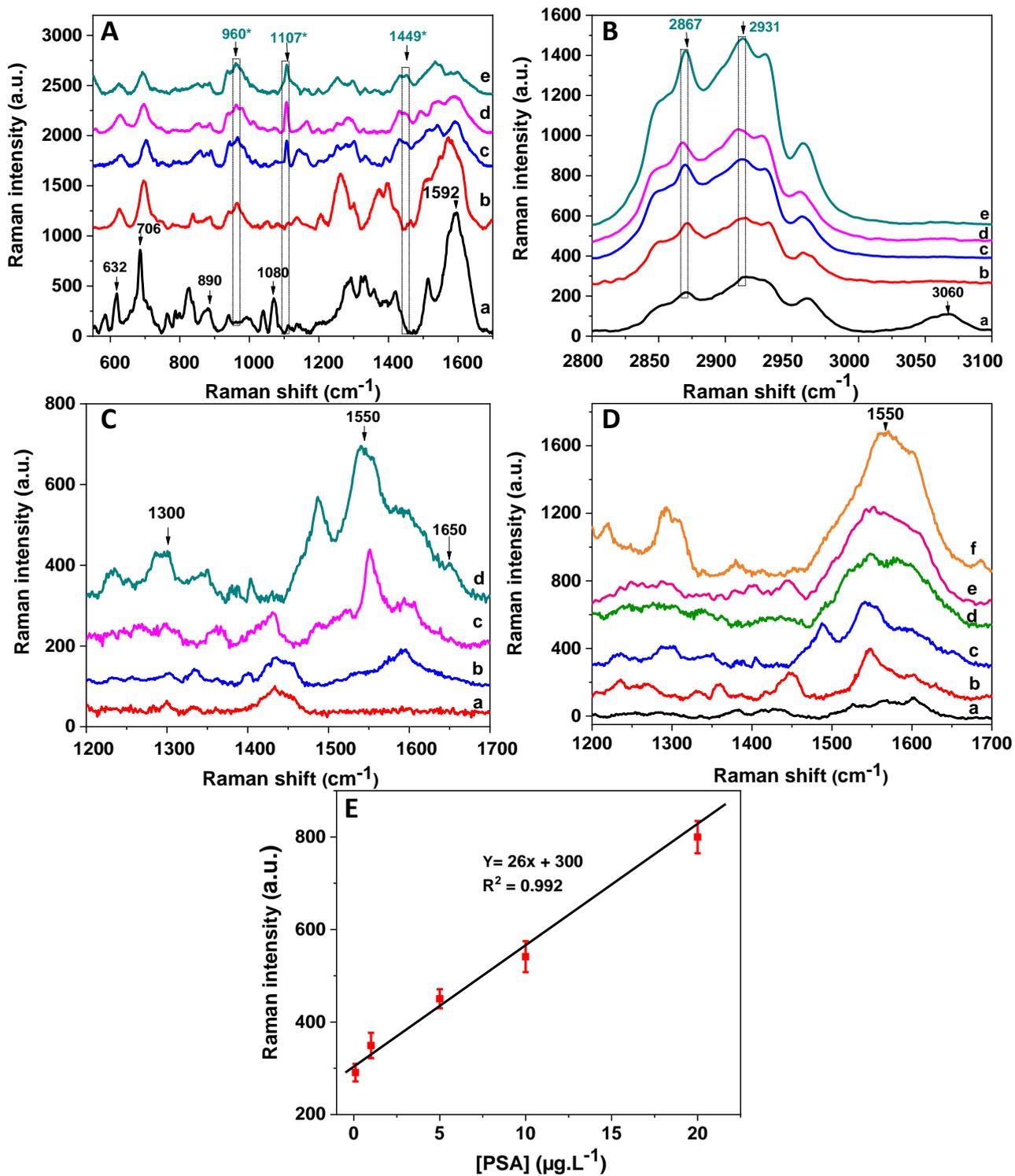


Figure 3. Raman spectra of MCH/AgNPs/SiNWs (a) and anti-PSA/MCH/AgNPs/SiNWs for different incubation times in aptamer ; 4h (b), 8h (c), 12h (d) and 16h (e) for two frequency ranges: 550-1700 cm^{-1} (**A**) and 2800-3100 cm^{-1} (**B**). Raman spectra of PSA(1 $\mu\text{g.L}^{-1}$)/anti-PSA/MCH/AgNPs/SiNWs for different incubation times ; 4h (a), 8h (b), 12h (c) and 16h (d) of the SERS substrate

(MCH/AgNPs/SiNWs) in aptamer solutions (C). SERS spectra of aptasensing system anti-PSA/AgNPs/SiNWs before (a) and after incubation in different PSA solutions with gradually increasing concentrations; 0.1 $\mu\text{g.L}^{-1}$ (b), 1 $\mu\text{g.L}^{-1}$ (c), 5 $\mu\text{g.L}^{-1}$ (d), 10 $\mu\text{g.L}^{-1}$ (e) and 20 $\mu\text{g.L}^{-1}$ (f) (D). Calibration curve of the developed aptasensor for PSA detection (E).

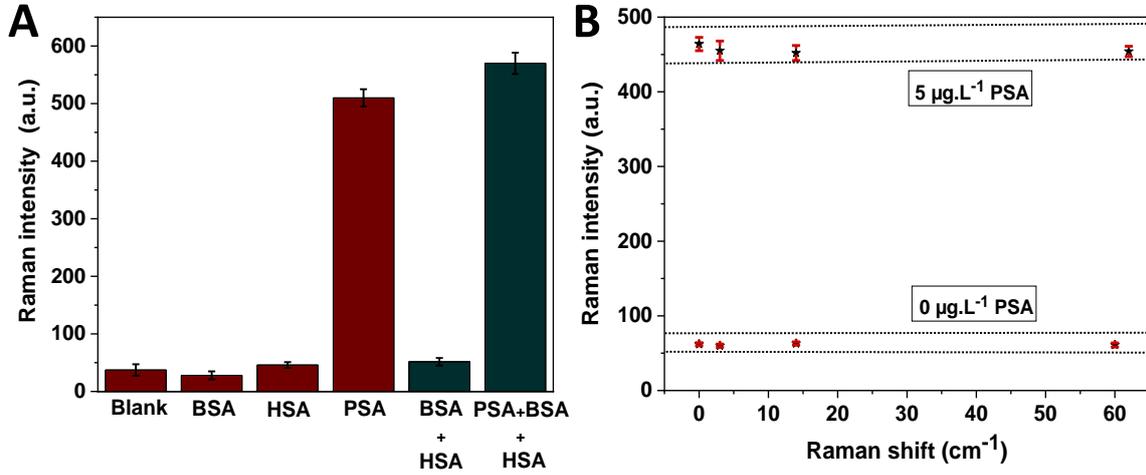


Figure 4. Analytical response of the aptasensor toward different proteins: PSA (10 $\mu\text{g.L}^{-1}$), BSA (10 $\mu\text{g.L}^{-1}$) and HSA (10 $\mu\text{g.L}^{-1}$) (A). Storage stability of the aptasensor stored for 62 days at 4 °C and its response in presence of 5 $\mu\text{g.L}^{-1}$ of PSA recorded periodically (B).

Table 1 : Comparison of the performances of some reported biosensors using electrochemical and optical technique with the present work.

Detection system	Method	Range of variation ($\mu\text{g.L}^{-1}$)	LOD ($\mu\text{g.L}^{-1}$)	Storage stability	Selectivity	Ref. (Year)
SPCE/GO-CO ₂ H/apta/cDNA/MB	DPV	0.001- 100	0.064×10 ⁻³	Two weeks	IgG, CEA and HSA	47 (2019)
MIP/PTBs/Au electrode	DPV	1-60	---	14 days	CEA and HSA	48 (2020)
Apt/MSF/Au	DPV	1-300	0.28	30 days	BSA, HIgG	20 (2017)
GO-AgNPs nanocomposite	LSV	5× 10 ⁻³ -20× 10 ⁻³	0.33× 10 ⁻³	----	CEA, AFP and BSA	49 (2019)

Au/apta-MIP	Capacitance	0.1- 100	0.001	-----	HK2 and HSA	50 (2016)
GCE/PANI/Au/Peptide	DPV	0.0001-100	0.085	7 days	CEA, IgG and HSA	32 (2019)
PS/PEG/COOH-PDOT	Fluorescence	3-15	2.05	---	CEA and AFP	51 (2018)
SERS dots/Ab ₁ /PSA/Ab ₂ /SERS dots	SERS	0.001-1000	0.11×10^{-3}	---	-----	52 (2016)
Glass/Au/A ₁ /PSA/Ab ₂ /Au-reporter	SERS	0.001-1000	4×10^{-3}	---	-----	53 (2003)
GO-AgNPs	Raman spectroscopy	0.0005-0.5	0.23	----	AFP, CEA, GSH, Lys, Ala, ALP, HSA and Glb	54 (2018)
anti-PSA/MCH/AgNPs/SiNWs	Raman spectroscopy	0.1-20	0.1	Two months	BSA and HSA	This work (2020)

SPCE: Screen-printed carbon electrodes, GO: graphene oxide, apta: anti-PSA aptamer, MB: methylene blue, DPV: differential pulse voltammetry, IgG: Immunoglobulin G, CEA: Carcino Embryonic Antigen, HSA: Human serum albumin, PTBs: poly[Toluidine blue], Au: gold electrode, MSF: Mesoporous silica thin films, BSA : Bovine serum albumin, AFP: Human alpha fetoprotein, HK2: Human glandular Kallikrein 2, MIP: molecularly imprinted polymer, GCE: glass carbon electrode, PANI: poly-aniline, PS-PEG-COOH: Polystyrene graft ethylene oxide, PDOT: polymer dots, AgNPs : silver nanoparticules, GSH: glutathione, Lys: lysine, Ala; alanine, ALP: alkaline phosphatase, GLB; globulin., Ab: antibody, LSV : linear sweep voltammetry.

Table 2 : Determination of PSA in spiked human blood serum

Samples	Serum (R)	Added (P) ($\mu\text{g.L}^{-1}$)	Found (F) ($\mu\text{g.L}^{-1}$)	RSD ² (%)	Recovery ³ (%)
1	ND ¹	3	3.07	5.0	97.7 ± 5
2	ND ¹	6	6.17	10.01	97.2 ± 10

¹ND: not detected, ²Relative standard deviation (RSD) of 3 measurements, ³Recovery (%) = $100 \times [R + P]/[F]$.